# PLA-Derived Lactate Esters as Plasma Precursors for Hydrophilic and Antibacterial Coatings

Monika Strašáková[1], Eva Domincová Bergerová[1*], Marián Lehocký[1], Veronika Hanuliak[1], Hana Pištěková[1], Michal Urbánek[1], Pavel Sťahel[2], Monika Stupavská[2], Jakub Klaban[1], Anežka Lengálová[3], Vladimír Sedlařík[1]

[1]Centre of Polymer Systems, Tomas Bata University in Zlin, tr. Tomase Bati 5678, 760 01 Zlin, Czech Republic

[2] Department of Plasma Physics and Technology, Faculty of Science, Masaryk University, Kotlarska 267/2, 611 37 Brno, Czech Republic

[3] Faculty of Humanities, Tomas Bata University in Zlin, Stefanikova 5670, 760 01 Zlin, Czech Republic

[*]Correspondence: Eva Domincová Bergerová, domincova_bergerova@utb.cz

## Abstract

This study introduces a novel concept of using lactate esters as unconventional, bio-based precursors for plasma-assisted deposition of multifunctional coatings. Unlike traditional plasma precursors, simple molecules such as methyl and ethyl lactate enable simultaneous enhancement of surface hydrophilicity and antibacterial activity on PET substrate. This work highlights a new pathway for designing advanced functional coatings using readily available ester-based compounds. Plasma deposition induced pronounced changes in surface chemistry and morphology through fragmentation–recombination mechanisms, leading to a significant increase in hydrophilicity, with the water contact angle decreasing from 76° to 27° for ethyl lactate and to 51° for methyl lactate. X-ray photoelectron spectroscopy confirmed the formation of oxygen- and nitrogen-containing functional groups, while the modified surfaces exhibited strong antibacterial performance, achieving up to 3.4 log reduction against Escherichia coli and 2.3 log reduction against Staphylococcus aureus (ISO 22196:2011). Crucially, this work establishes lactate esters as a new and versatile class of plasma precursors, demonstrating that waste-derived, low-molecular-weight compounds can outperform conventional systems in simultaneously imparting hydrophilicity and

antimicrobial functionality, thereby redefining their role in plasma processing and providing a scalable platform for multifunctional coatings within a circular economy framework.

**Keywords:** Plasma deposition, surface modification, antibacterial properties, polylactic acid ester, polymeric material

## 1. Introduction

Plastics have become some of the most widely produced and utilized materials globally. The substantial growth in production and consumption has led to the accumulation of large quantities of waste, presenting a significant challenge for environmentally responsible disposal. Projections suggest that annual global production will result in approximately 12,000 Mt of waste by 2050. As production continues to rise, the resulting excess waste poses an increasing environmental threat to both humanity and the planet. Traditional waste management methods, such as landfilling, are inadequate due to the non-biodegradable nature of many conventional polymers, which leads to their prolonged persistence in disposal sites [1,2].

In response to this issue, there has been growing interest in alternative materials that retain the functional properties of conventional materials while being more environmentally sustainable. This has spurred the development of biodegradable polymers, now used in the production of eco-friendly alternatives. Bio-based polymers, aligned with the principles of sustainable production and consumption, have emerged as key materials in this context [1,3]. The demand for bioplastics has surged as a viable alternative to traditional polymers, primarily due to their renewable nature, non-toxicity, biodegradability, and biocompatibility. Furthermore, modern bio-based materials exhibit mechanical properties that are comparable to those of their fossil-derived counterparts [4].

Among the various bioplastics, polylactic acid (PLA) is considered one of the most promising candidates for replacing non-degradable polymers such as polyethylene terephthalate (PET) and polystyrene (PS), particularly in food packaging applications. However, the increasing production and use of PLA, combined with inadequate consumer waste collection systems and limited technological solutions for its effective recycling, have resulted in frequent cross-contamination of recycling streams. In practice, PLA waste is often unintentionally mixed with conventional plastic waste, most notably with PET recycling streams, where even small

amounts of PLA can negatively affect the recycling process and the quality of recycled PET. Consequently, the simultaneous management of biodegradable and non-biodegradable plastics within the same waste management infrastructure remains a significant challenge [5,6].

A promising approach to this issue is processing waste of these materials, which involves a systematic technological approach and the repurposing of waste. For PLA, potential processing waste routes include landfilling or composting, incineration, and both mechanical and chemical recycling. However, these methods pose several challenges, including the loss of valuable polyhydroxy acids, degradation of soil quality, and the release of toxic exhaust gases.

In contrast, chemical recycling represents a promising strategy for extending the lifecycle of post-consumer plastic waste. In the case of PLA, chemical recycling is considered an economically viable and environmentally sustainable approach, as it enables the depolymerization of the ester backbone into valuable low-molecular-weight compounds, primarily lactate esters, which can be further utilized as functional precursors in various applications [3]. An important advancement in surface engineering is plasma polymerization, a process in which reactive species generated in the plasma phase - such as radicals, ions, and excited molecules - are formed from a gaseous precursor and subsequently deposited onto a substrate, where they recombine to form a highly cross-linked polymer-like thin film [7]. Plasma-based technologies offer several advantages, including solvent-free processing, low thermal load, and reduced use of hazardous chemicals. Importantly, plasma treatment modifies only the surface of a material while preserving its bulk properties. In addition, plasma exposure can introduce polar functional groups, such as hydroxyl and carboxyl moieties, which significantly enhance surface wettability, as evidenced by the reduction in water contact angle following treatment [8].

However, further research is needed on the plasma processing of PLA esters, such as ethyl lactate and methyl lactate, which are decomposition products resulting from chemical recycling. Furthermore, existing literature largely focuses on the combination of commercially available ethyl lactate with additives, nanoparticles, or composites, rather than exploring plasma deposition or polymerization of PLA decomposition products [9–11].

This study investigates the application of plasma technology in the processing of products derived from the chemical recycling of biopolymers, with a focus on optimizing ecological sustainability. By integrating these approaches, this work not only offers a sustainable

solution for addressing the projected accumulation of plastic waste by 2050 but also produces materials with enhanced properties suitable for a wide range of applications, marking a significant step forward in the implementation of circular economy practices and the development of sustainable materials.

Considering the need for more sustainable valorization routes for chemically recycled PLA monomers and the scarcity of studies on their plasma-assisted deposition, there is a clear research gap. Moreover, PET is one of the most common non-biodegradable packaging materials and provides an ideal substrate for surface modification, where tailored coatings can impart new functionalities without altering bulk properties.

Therefore, the aim of this study is to investigate the plasma-assisted deposition of methyl lactate and ethyl lactate obtained through chemical recycling of PLA onto PET substrates under atmospheric pressure conditions. The work focuses on (i) characterizing the chemical composition and morphology of the deposited layers, (ii) evaluating their hydrophilic properties, and (iii) assessing their antibacterial performance. By linking PLA waste chemical recycling with plasma deposition, this work establishes a link between chemical recycling of biopolymers and plasma surface engineering by demonstrating that methyl and ethyl lactate obtained from PLA can serve as effective precursors for the preparation of hydrophilic and antibacterial coatings on PET substrates.

## 2. Materials and methods

### 2.1 Materials

Polyethylene terephthalate (PET) films (TFP universal a.s., Dobřejovice, Czech Republic) with dimensions of 120 mm × 1000 mm and a thickness of 1 mm were used as substrates for deposition and served as a reference material. The monomers utilized for plasma deposition were ethyl lactate and methyl lactate, derived from virgin PLA (Ingeo™ 2003D, NatureWorks®, Minneapolis, USA), specifically prepared through the solvolysis process and their purity was determination as described by Domincova Bergerova et al. [12]. The bacterial strains *Staphylococcus aureus* (CCM 4516) and *Escherichia coli* (CCM 4517) were sourced from the Czech Collection of Microorganisms, Masaryk University (Brno, Czech Republic).

### 2.2 Sample preparation - plasma deposition

Plasma deposition was conducted according to Šťahel et al. [13], with a few minor adjustments. PET films were prepared and served as the initial substrates for plasma deposition. Thin film plasma polymerization was performed in a specially designed reactor

equipped with a surface dielectric barrier discharge (SDBD) system. The reactor comprised three primary components: an electrode system, a cooling system, and a high-voltage generator. The complete deposition apparatus was situated within a fume hood throughout the deposition procedure, and the resultant gaseous products were continuously evacuated. The SDBD electrode setup consisted of 11 upper rotating cylindrical brass electrodes with a diameter of 1 cm and a length of 10.4 cm, spaced 2 mm apart. Below the cylindrical electrodes, a flat lower electrode of the size of 13.6 x 10 cm was positioned and covered with a 1 mm thick mica dielectric. The substrate was periodically moved along the upper and lower electrodes at a speed of 3 cm/min. As the upper electrodes rotated, they contacted the moving substrate. The lower electrode was grounded, whereas the upper electrodes were connected to a high-voltage AC power supply (Lifetech, Brno, Czech Republic) using a sinusoidal voltage with an amplitude of 11 kV and a frequency of 12 kHz. The power input to the system was set to 150 W. A schematic of the experimental setup is presented in Fig. 1.

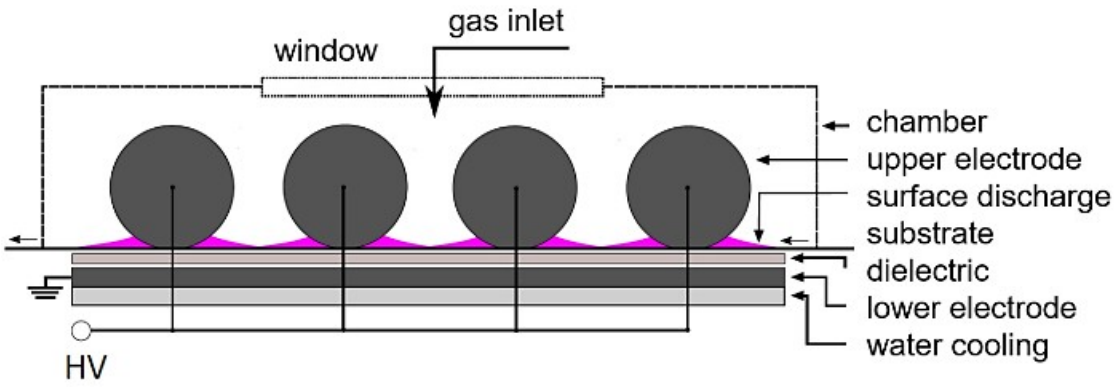



Figure 1: Schematic of the deposition setup [13]

As the working gas, nitrogen was used; it was supplied between the upper electrodes at a flow rate of 3 slm. The monomers ethyl lactate and methyl lactate as precursors were introduced into the nitrogen flow at rates ranging from 250 to 1000 sccm. The substrate passed through the discharge three times, with a total deposition time of 6.5 seconds. Throughout the deposition process, the apparatus was water-cooled to maintain a consistent electrode temperature.

## 2.3 Characterization of chemical structure

### 2.3.1 Fourier transform infrared spectroscopy with attenuated total reflection

Fourier transform infrared spectroscopy with attenuated total reflection (ATR-FTIR) spectra were recorded for all polymer films using a Nicolet iS5 spectrometer (Thermo Fisher

Scientific, Waltham, USA), equipped with a Ge crystal. The analysis was conducted over a range of 600–4000 $cm^{-1}$ with a resolution of 4 $cm^{-1}$, and 64 scans were averaged per spectrum. Data processing was performed using OMNIC software (Thermo Fisher Scientific, Waltham, USA).

#### 2.3.2 X-ray photoelectron spectroscopy

X-ray photoelectron spectroscopy (XPS) analysis was conducted using an ESCALAB 250Xi system (Thermo Fisher Scientific, East Grinstead, UK), utilizing an X-ray beam with a power output of 200 W and a spot size of 650 microns. Survey spectra were collected with a pass energy of 50 eV and an energy step of 1 eV, while high-resolution scans were performed with a pass energy of 20 eV and an energy step of 0.1 eV. To mitigate surface charging effects, an electron flood gun was employed. All spectra were referenced to the C1s hydrocarbon component with a binding energy of 284.8 eV. Data calibration, processing, and fitting were performed with Avantage software (version 5.9925).

### 2.4 Characterization of hydrophilic properties

The wettability of the sample surface was evaluated by measuring the static contact angle (CA) using a SEE System instrument (Advex Instruments, Czech Republic), equipped with a UVC camera and a high-resolution glass objective lens. Three test liquids were employed: ultrapure water (resistivity 18.2 MΩ·cm, Milli-Q, Merck), diiodomethane (≥99%, Sigma Aldrich), and ethylene glycol (≥99%, Sigma Aldrich). Droplets of 3 µL were deposited on the surface using a micropipette, and images were acquired approximately 1 s after deposition. For each liquid and sample, five independent measurements were performed, and the average values are reported. The images were analyzed using the SEE System software to determine the contact angle. The total surface free energy (γ^tot) and its dispersive (γ^d) and polar (γ^p) components were calculated according to the **Owens–Wendt–Rabel–Kaelble (OWRK) method** [14], using the following relationship (1):

$$\gamma_L(1+\cos\theta) = 2\left(\sqrt{\gamma_S^d \gamma_L^d} + \sqrt{\gamma_S^p \gamma_L^p}\right) \quad (1)$$

where $\gamma_L$ and $\gamma_S$ are the liquid and solid surface tensions, with dispersive (d) and polar (p) contributions.

### 2.5 Characterization of morphology

#### 2.5.1 Scanning electron microscopy

Scanning electron microscopy (SEM) images of the sample surfaces and fracture areas were obtained using a Nova NanoSEM 450 (FEI, USA). Prior to imaging, the samples were coated with a gold–palladium alloy to improve electrical conductivity; the coating was deposited by sputtering, which lasted 60 s and resulted in the film thickness of approximately 10 nm, with a grain size in the range of 3–5 nm. The SEM measurements were conducted at a working distance (WD) of 4.7 mm and an accelerating voltage (HV) of 5 kV.

### 2.5.2 Atomic force microscopy

The surface topography of treated and untreated PET films was characterized using an atomic force microscope (AFM) model Dimension-Icon. Measurements were conducted at a scan rate of 1 Hz with a resolution of 256 × 256 pixels in tapping mode under ambient air conditions. For the measurements, a silicon-nitride probe ScanAsyst-Air (both Bruker, Billerica, MA, USA) with a resonant frequency of 70 kHz and a stiffness constant of 0.4 $N \cdot m^{-1}$ was employed.

## 2.6 Characterization of antibacterial properties

The antibacterial properties of the samples were evaluated following the ISO 22196:2011 standard. Prior to testing, the samples were disinfected with 70% denatured ethanol. Bacterial suspensions of *E. coli* ($3.1 \times 10^5$ CFU/mL) and *S. aureus* ($4.0 \times 10^5$ CFU/mL) were prepared in 1/500 nutrient broth (HiMedia laboratories, Mumbai, India). A 100 µL aliquot of bacterial suspension was applied to the surface of the samples (25 × 25 mm), which were then covered with polypropylene foil (20 × 20 mm). Afterwards, the samples were incubated at 35 °C and 100% relative humidity for 24 hours. Following incubation, the polypropylene foil was removed, and the samples were washed with soya casein digest lecithin polysorbate broth, which was subsequently collected.

The antibacterial activity was evaluated using equation (2):

$$R = U_t - A_t \qquad (2)$$

where $R$ is the antibacterial activity, $U_t$ means the common logarithm of the viable bacteria count (CFU $cm^{-2}$) recovered from the untreated (blank) sample, and $A_t$ stands for the count for the treated specimen after 24 hours [15].

For subsequent analyses, the plasma-treated samples at a flow rate of 1000 sccm were used.

## 3 Results and discussion

### 3.1 Characterization of chemical structure

#### 3.1.1 ATR-FTIR Analysis

As seen in Figure 2, the ATR-FTIR spectra of all samples, including methyl lactate/PET, ethyl lactate/PET, and bare PET, exhibit near-identical profiles. Comparison of the spectra using the OMNIC software library revealed a greater than 99% match for all samples. Notably, the ethyl lactate/PET and the methyl lactate/PET samples exhibited changes in the region between 3500 and 2600 $cm^{-1}$, corresponding to O-H or N-H bonds, which may contribute to increased hydrophilicity [16]. The spectral similarity suggests that the deposited plasma films are relatively thin, with the FTIR signal predominantly influenced by the underlying PET substrate. Moreover, as no previous studies have reported FTIR spectra for plasma-treated ethyl lactate and methyl lactate, this aspect remains insufficiently understood and warrants further systematic investigation.

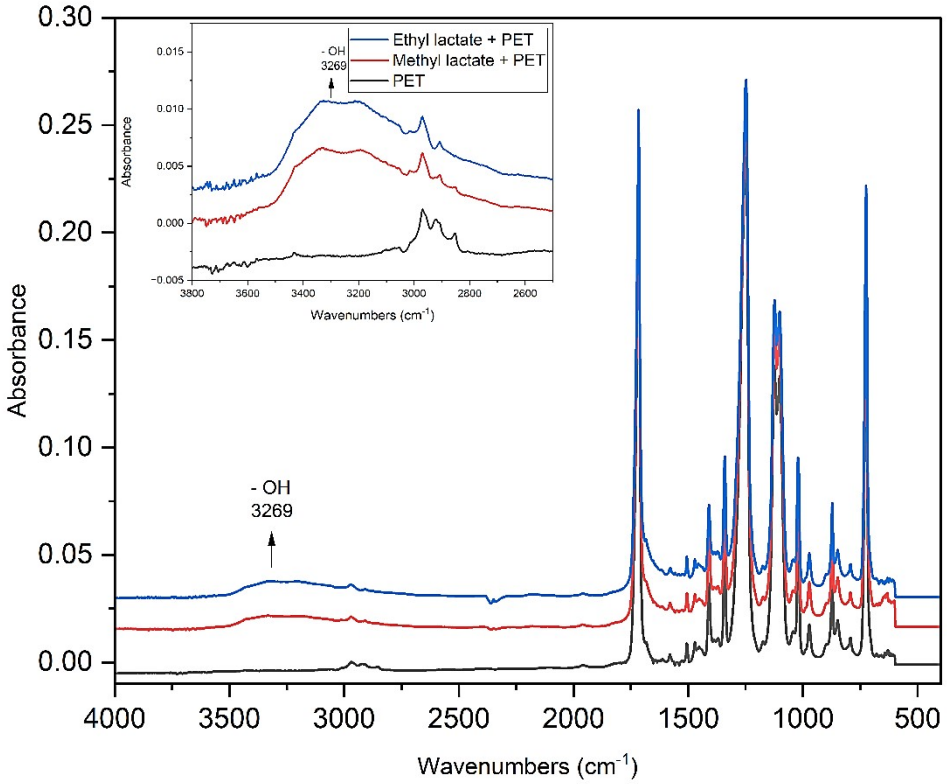



Figure 2: FTIR spectra of samples: methyl lactate/PET (flow rate 1000 sccm), ethyl lactate/PET (flow rate 1000 sccm) and bare PET.

### 3.1.2 Surface composition of samples

The surface composition of samples was followed with X-ray photoelectron spectroscopy and the results are summarized in Table 1. As can be seen, the reference PET sample exhibits 74% carbon and 24% oxygen, which is consistent with theoretical expectations [17]. The O/C ratio, indicative of increased surface oxygen-containing functionalities, is 0.324. For the plasma-modified methyl lactate/PET sample, the surface consists of 61% carbon, 27% oxygen, and 11% nitrogen, resulting in an increased O/C ratio of 0.443. In the case of the ethyl lactate/PET sample, nitrogen content was markedly higher reaching 25%, with oxygen also present at 25%, and carbon at 50%. The O/C ratio for this sample is 0.5, representing an over 1.5-fold increase compared to the reference PET sample. These results indicate substantial surface functionalization following plasma deposition.

Table 1 Sample elemental surface composition (%) and bond distributions (%) according to XPS analysis. Samples: methyl lactate/PET (flow rate 1000 sccm), ethyl lactate/PET (flow rate 1000 sccm) and bare PET.

| | | **PET reference (%)** | **methyl lactate/PET (%)** | **ethyl lactate/PET (%)** |
|---|---|---|---|---|
| | **C** | 74 | 61 | 50 |
| | **O** | 24 | 27 | 25 |
| | **N** | 0 | 11 | 25 |
| **C1s** | **C=C** | 15 | 11 | 6 |
| | **C-C/C-H** | 50 | 45 | 28 |
| | **C-N** | 0 | 8 | 13 |
| | **C-O** | 23 | 13 | 10 |
| | **C=O** | 0 | 2 | 8 |
| | **N-C=O** | 0 | 3 | 10 |
| | **COO** | 12 | 18 | 25 |
| **N1s** | **-C=N** | - | 5 | 5 |
| | **-C-NH-** | - | 69 | 57 |
| | **-CONH-** | - | 16 | 30 |
| | **nitrate** | - | 10 | 8 |

For a more detailed analysis of the effect of plasma modification, high-resolution XPS spectra of the C1s and N1s were measured (Figure 3). The figure reveals distinct differences in surface chemistry between the reference PET and the plasma-modified samples, with variations also observed between the methyl and ethyl lactate treatments. Deconvolution of these spectra provided further insights into the formation of different functional groups.

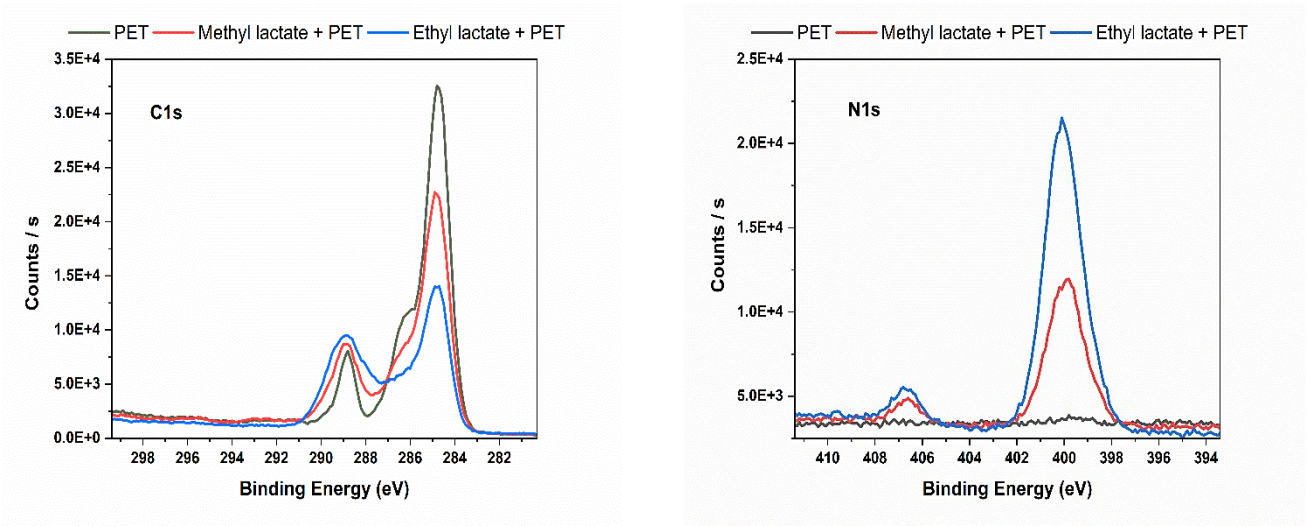



Figure 3: Overlay of XPS C1s and N1s spectra of the PET reference and plasma modified samples (methyl lactate/PET and ethyl lactate/PET).

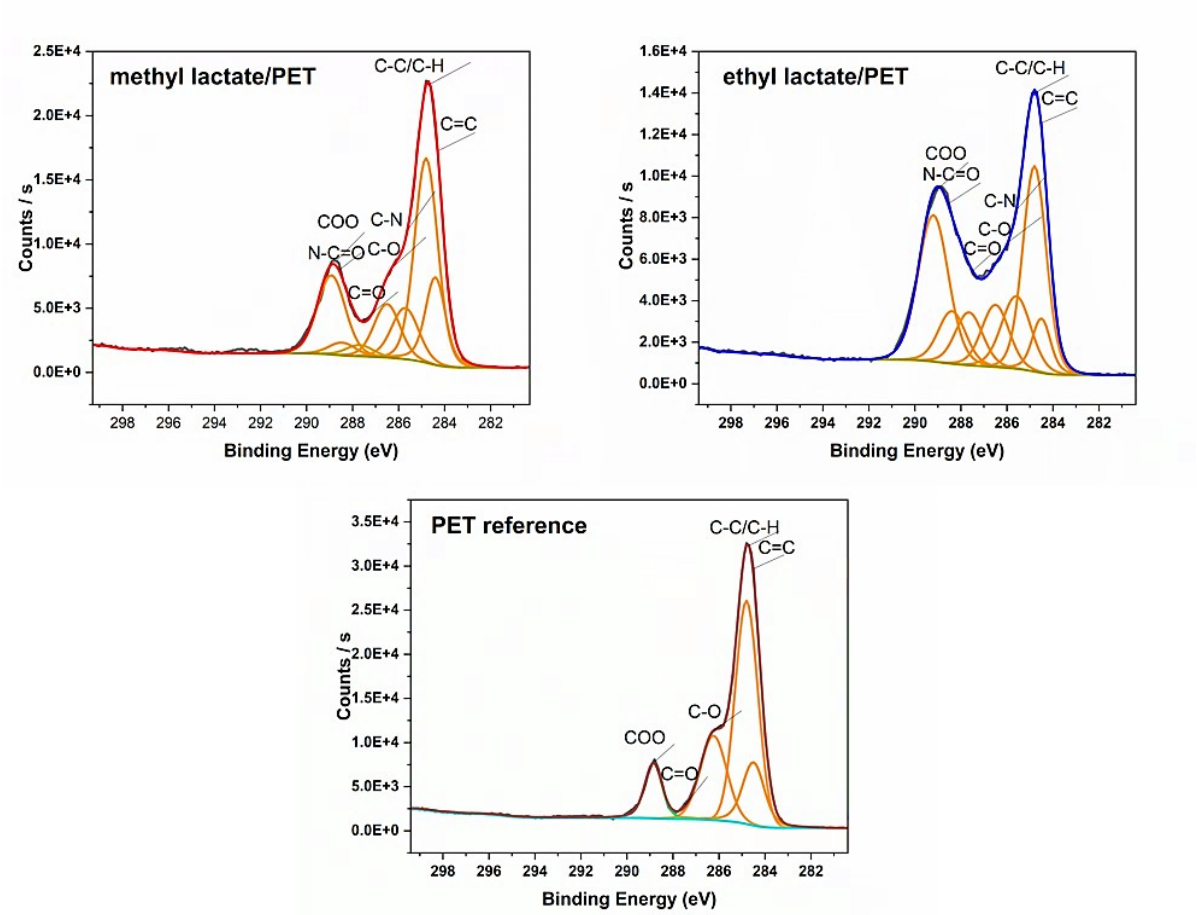
The C1s spectra of the reference PET sample were deconvoluted into four distinct peaks with binding energies of 284.5 eV, 284.8 eV, 286.4 eV, and 289 eV, corresponding to carbon atoms in C=C, C–C/C–H, C–O, and COO functional groups, respectively. Following plasma modification, three additional peaks emerged at binding energies of 285.7 eV, 287.7 eV, and 288.4 eV, which are assigned to C–N, C=O, and N–C=O functional groups, respectively (Figure 4) [18],[19], [20],[21],[22] . The relative concentrations of all detected carbon functional groups on the surface of the analyzed samples are presented above in Table 1.

Deconvolution of the C1s spectra reveals a progressive reduction in the relative contributions of nonpolar carbon environments (C=C and C–C/C–H) compared to the reference PET. The fraction of C=C bonds decreases from 15% in pristine PET to 11% in methyl lactate/PET and to 6% in ethyl lactate/PET, while C–C/C–H bonds drop from 50% to 45% and 28%,

respectively. Importantly, this decrease does not indicate chemical consumption or transformation of the aromatic structures of PET. Rather, it reflects attenuation of the XPS signal from the underlying polymer due to coverage by a thin plasma-deposited film. Because XPS probes only the outermost nanometers of the surface, the detected bonding environment primarily represents the newly formed coating rather than the bulk PET substrate. Concurrently, there is a pronounced increase in polar oxygen- and nitrogen-containing functional groups. Signals assigned to C–N, C=O, N–C=O, and COO groups emerge or increase significantly after plasma deposition. The COO contribution rises from 12% in the reference PET to 18% in methyl lactate/PET and 25% in ethyl lactate/PET, while C–N and N–C=O functionalities are absent in pristine PET but clearly present in both modified samples. These changes confirm that plasma-assisted deposition introduces a highly functionalized surface layer enriched in ester, carbonyl, amide, and amine-related structures, which are expected to increase surface polarity and hydrophilicity.

The N1s spectra further support this interpretation. The plasma-treated samples display peaks corresponding to imine (-C=N, 398.6 eV), amine (-C-NH-, 400 eV), amide (-CONH-, 401.0 eV), and nitrate ($NO_3^-$, 406.7 eV) functionalities (Figure 5). Nitrogen is incorporated predominantly in the form of amine groups, which represent the major nitrogen species in both ester-derived coatings. In addition, amide and imine functionalities are detected, together with minor contributions attributed to nitrate species. The ethyl lactate/PET sample shows a higher fraction of amide groups and an overall nitrogen content approximately twice that of the methyl lactate/PET sample, indicating more extensive nitrogen-containing functionalization.

Overall, the XPS results confirm successful plasma-assisted deposition of ester-derived coatings onto PET. The observed spectral changes are governed by the formation of a new, thin, heteroatom-rich surface layer that masks the underlying PET signal, rather than by chemical modification of the PET backbone itself. Among the two precursors, ethyl lactate leads to a more strongly functionalized surface, characterized by higher nitrogen incorporation and a greater abundance of carbonyl- and amide-related groups. These surface chemical changes are consistent with the observed enhancement in hydrophilicity and are likely to contribute to the functional performance of the coatings.

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294 Figure 4: High-resolution XPS C1s spectra of the PET reference and plasma-modified
295 methyl lactate/PET and ethyl lactate/PET samples.

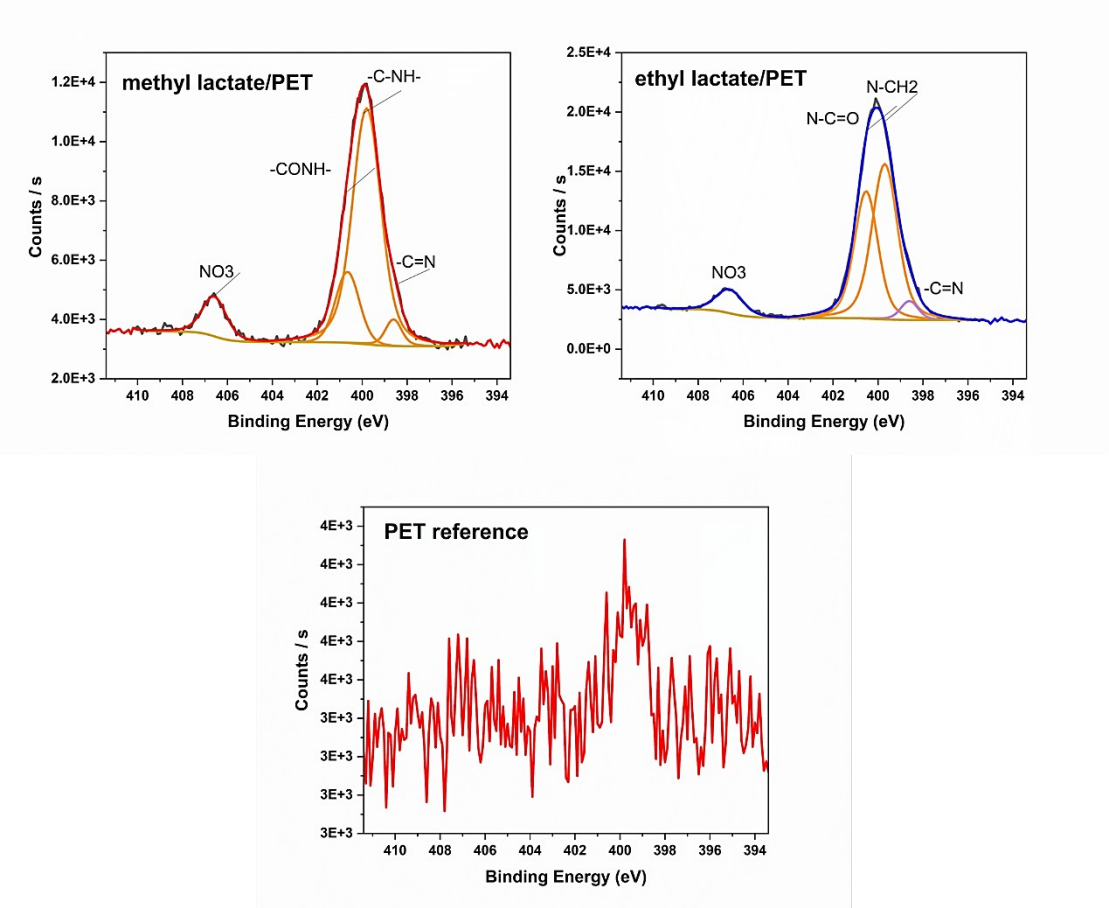


Figure 5: High-resolution XPS N1s spectra of the PET reference and plasma-modified methyl lactate/PET and ethyl lactate/PET samples.

## 3.2 Hydrophilicity of materials - contact angle and surface energy evaluation

The contact angle (CA) values and calculated surface free energy (SFE) of the samples are summarized in Table 2. Water CA values serve as a qualitative indicator of hydrophilic versus hydrophobic character, whereas the complete set of contact angles obtained with different probe liquids is required for quantitative SFE analysis.

As shown in Table 2, plasma modification progressively enhanced the hydrophilic properties of both ethyl lactate and methyl lactate coatings. For ethyl lactate, the most pronounced hydrophilicity was observed at a monomer flow rate of 1000 sccm, with a water CA of 27 ± 2°. In contrast, the lowest CA achieved for methyl lactate under the same conditions was 51 ± 3°. These results indicate that ethyl lactate plasma layers introduce a higher density of polar functionalities than methyl lactate, consistent with XPS findings that revealed greater incorporation of oxygen- and nitrogen-containing groups. Such a pronounced difference in wettability is particularly relevant, as previous studies have shown that only surfaces with

very low water contact angles (below ~30°) are capable of sustaining stable hydration layers that effectively alter interfacial interactions [23,24]. The evolution of water CA values for both coatings is presented graphically in Figure 6.

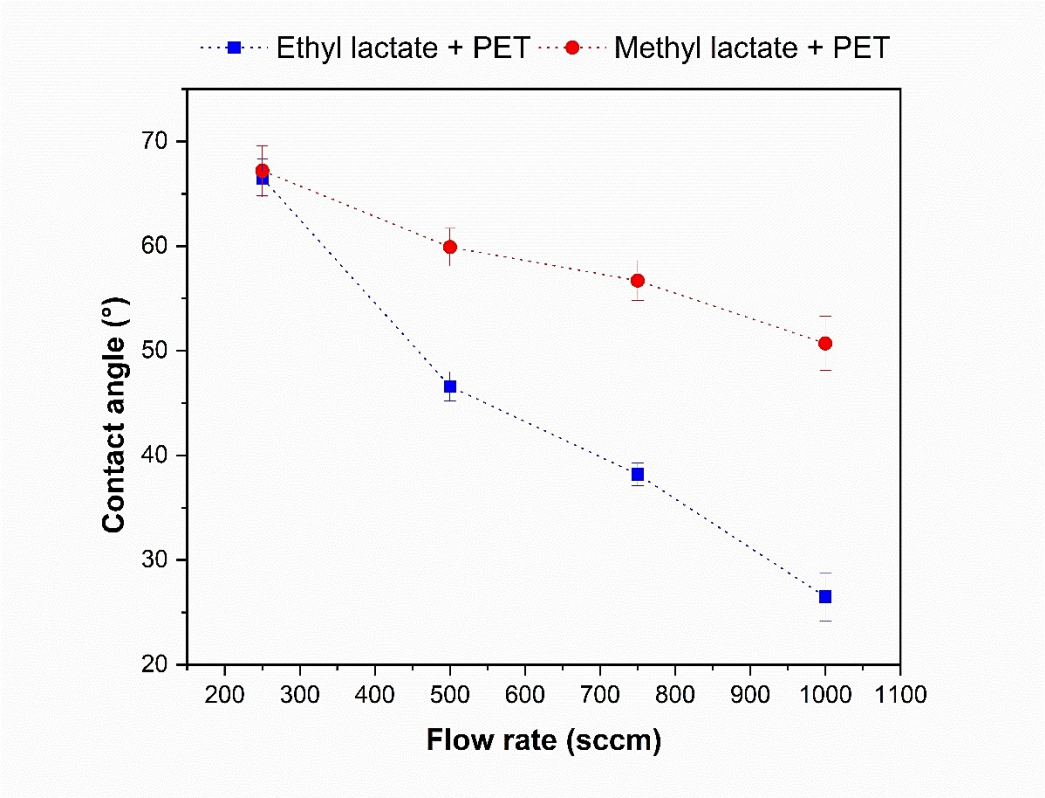


Figure 6: The water contact angle values for plasma-polymerized ethyl lactate/PET and methyl lactate/PET films (RSD = 3 ± 1).

The Owens–Wendt analysis further confirmed that plasma treatment predominantly increased the polar contribution (γ^p) of SFE, while the dispersive part (γ^d) remained nearly unchanged. For example, ethyl lactate at 1000 sccm exhibited a total SFE of 60 mJ·m$^{-2}$ with a dominant polar fraction, compared to only 46 mJ·m$^{-2}$ for untreated PET. This trend is supported by XPS (Table 1), which detected incorporation of polar oxygen- and nitrogen-containing groups (–OH, –C=O, –C–N, N–C=O). Importantly, FTIR spectroscopy (Figure 2) provided complementary evidence, revealing a more pronounced presence of –OH functionalities in ethyl lactate compared to methyl lactate coatings. In sum, the XPS and FTIR results converge on the conclusion that plasma deposition promotes the incorporation of hydroxyl and nitrogen-containing groups, which are directly responsible for the enhanced surface polarity and hydrophilicity.

Comparable trends have been documented for other plasma-modified polymers. Lai et al. demonstrated that plasma treatment of polymers results in a significant decrease in water CA and a corresponding increase in SFE, primarily due to oxygen incorporation [25]. Johnston and Ratner reported that plasma-polymerized organic films exhibit a marked increase in γ^p following the introduction of heteroatom-containing moieties [26], while Cui and Brown showed that in plasma-treated polypropylene the rise in SFE is attributable predominantly to γ^p [27]. More recent work corroborates these findings, including the deposition of nanocomposite coatings from ethyl lactate [27] and rapid hydrophilization via atmospheric plasma polymerization [28].

Analogous effects of plasma treatment on the enhancement of surface polarity and wettability have also been reported for polycarbonate [29], PET films modified by plasma-based ion implantation [30], and PET track membranes [31], underlining the universal nature of plasma-induced hydrophilization across diverse polymeric systems.

Overall, both the present data and the literature consistently indicate that the hydrophilization of PET surfaces coated with plasma-polymerized lactate esters originates from the introduction of polar functional groups, particularly hydroxyl and nitrogen species, leading to an increase in γ^p that clearly outweighs changes in dispersive interactions.

Table 2 Contact angles of plasma-polymerized films deposited at different monomer flow rates. The PET reference represents the untreated substrate.

| **Sample** | **Flow Rate (sccm)** | **Contact Angle (°)** | | | **Surface Free Energy ($mJ/m^2$)** |
|---|---|---|---|---|---|
| | | Water | $CH_2I_2$ | Ethylene glycol | Total |
| 0. PET reference | / | 76 ± 1 | 27 ± 1 | 31 ± 1 | 46 |
| 1. ethyl lactate/PET | 250 | 67 ± 2 | 32 ± 1 | 36 ± 1 | 48 |
| 2. ethyl lactate/PET | 500 | 47 ± 1 | 33 ± 2 | 29 ± 1 | 55 |
| 3. ethyl lactate/PET | 750 | 38 ± 1 | 33 ± 2 | 25 ± 0 | 55 |
| 4. ethyl lactate/PET | 1000 | 27 ± 2 | 46 ± 2 | 29 ± 2 | 60 |

| | | | | | |
|---|---|---|---|---|---|
| 5. methyl lactate/PET | 250 | 67 ± 2 | 30 ± 3 | 27 ± 3 | 50 |
| 6. methyl lactate/PET | 500 | 60 ± 2 | 32 ± 1 | 36 ± 2 | 50 |
| 7. methyl lactate/PET | 750 | 57 ± 2 | 37 ± 2 | 23 ± 2 | 51 |
| 8. methyl lactate/PET | 1000 | 51 ± 3 | 48 ± 2 | 48 ± 1 | 50 |

## 3.3 Morphology of the tested materials

### 3.3.1 Scanning electron microscopy

Figure 7 illustrates the surface morphology of PET in response to plasma treatment. Figure 7A presents a typical image of untreated PET film, characterized predominantly by the presence of irregular protrusions on the surface. In contrast, plasma-treated samples, depicted in Figures 7B and 7C, exhibit notable modifications in surface topography. Although some irregular protrusions remain visible following plasma treatment, the overall surface appears considerably smoother. Moreover, Figure 7C demonstrates an even greater degree of surface smoothness compared to the untreated PET, with only a few residual protrusions observed.

These findings suggest that a substantial number of micrometer-scale protrusions are effectively reduced or eliminated, a phenomenon that can be primarily attributed to the bombardment of the sample surface by energetic particles generated during the plasma treatment process [25].

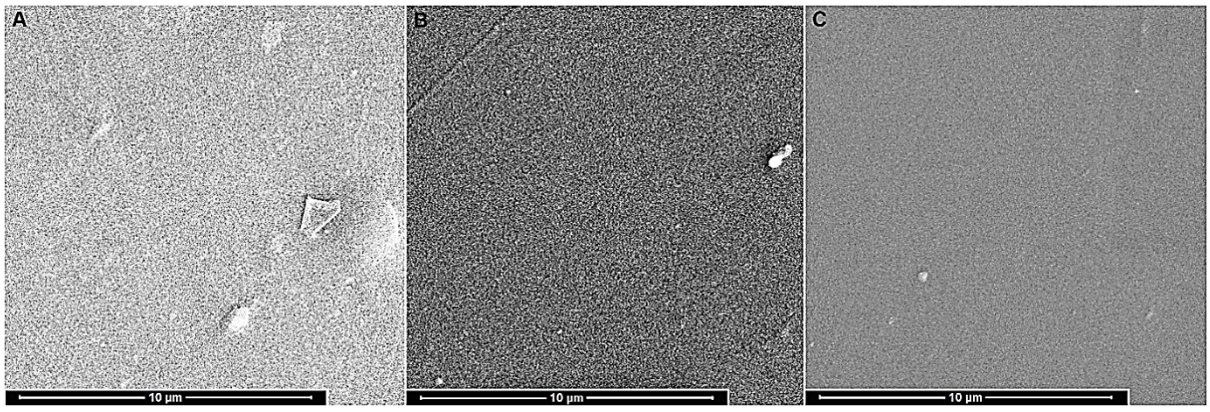


Figure 7: Representative SEM images of PET sample surfaces, obtained at approximately 10,000× magnification (scale bar: 10 µm), illustrating surface morphology variations: (A) untreated PET film, (B) ethyl lactate plasma-treated sample (nitrogen flow rate: 1000 sccm), and (C) methyl lactate plasma-treated sample (nitrogen flow rate: 1000 sccm).

### 3.3.2 Atomic force microscopy

Figure 8 presents atomic force microscopy (AFM) images illustrating the surface topography of samples. The surface topography of a reference sample, untreated polyethylene terephthalate (PET) film, was analyzed using AFM in tapping mode. Similarly, the surface morphology of samples coated with plasma-prepared layers of methyl lactate/PET and ethyl lactate/PET was examined under identical conditions. The scanned surface area measured 2 × 2 µm. The images on the left part of the figure depict three-dimensional representations of the individual samples, while the corresponding two-dimensional models are displayed on the right.

The untreated PET surface exhibits a heterogeneous morphology with relatively pronounced nanoscale undulations and irregular height fluctuations reaching approximately 60 nm. This morphology is typical for semi-crystalline PET films and reflects the coexistence of amorphous and crystalline domains formed during film processing. The root mean square roughness of pristine PET (Rq ≈ 2.6 nm) indicates a comparatively heterogeneous surface with locally exposed microstructural features [32].

Following plasma deposition, both modified samples show a substantial transformation of the original PET topography, confirming the formation of a continuous plasma-polymerized layer. The ethyl lactate/PET surface is characterized by a fine granular nanostructure composed of uniformly distributed nanoscale features, with height variations on the order of several nanometers and a reduced roughness (Rq ≈ 1.6 nm). This observation aligns with the findings of Milaniak et al., who reported that plasma polymerization of ethyl lactate results in a less pronounced surface roughness [33]. In contrast, the methyl lactate/PET sample displays a more homogeneous and smoother morphology with smaller, more compact nanoscale features, also exhibiting height variations on the order of several nanometers and a further reduction in roughness (Rq ≈ 0.6 nm). The smoother morphology suggests the formation of a denser and more uniform plasma polymer layer, which may be attributed to differences in precursor fragmentation pathways and polymerization kinetics. Similar morphology evolution has been reported for plasma polymer films derived from oxygen-containing organic precursors, where film densification and uniform growth lead to reduced nanoscale roughness [34,35]

In conclusion, from a functional perspective, the nanoscale roughness values of all plasma-modified samples remain in the low-roughness regime. The combination of reduced

roughness and increased surface polarity promotes the formation of stable hydration layers, which are known to influence wettability and bacterial adhesion [36].

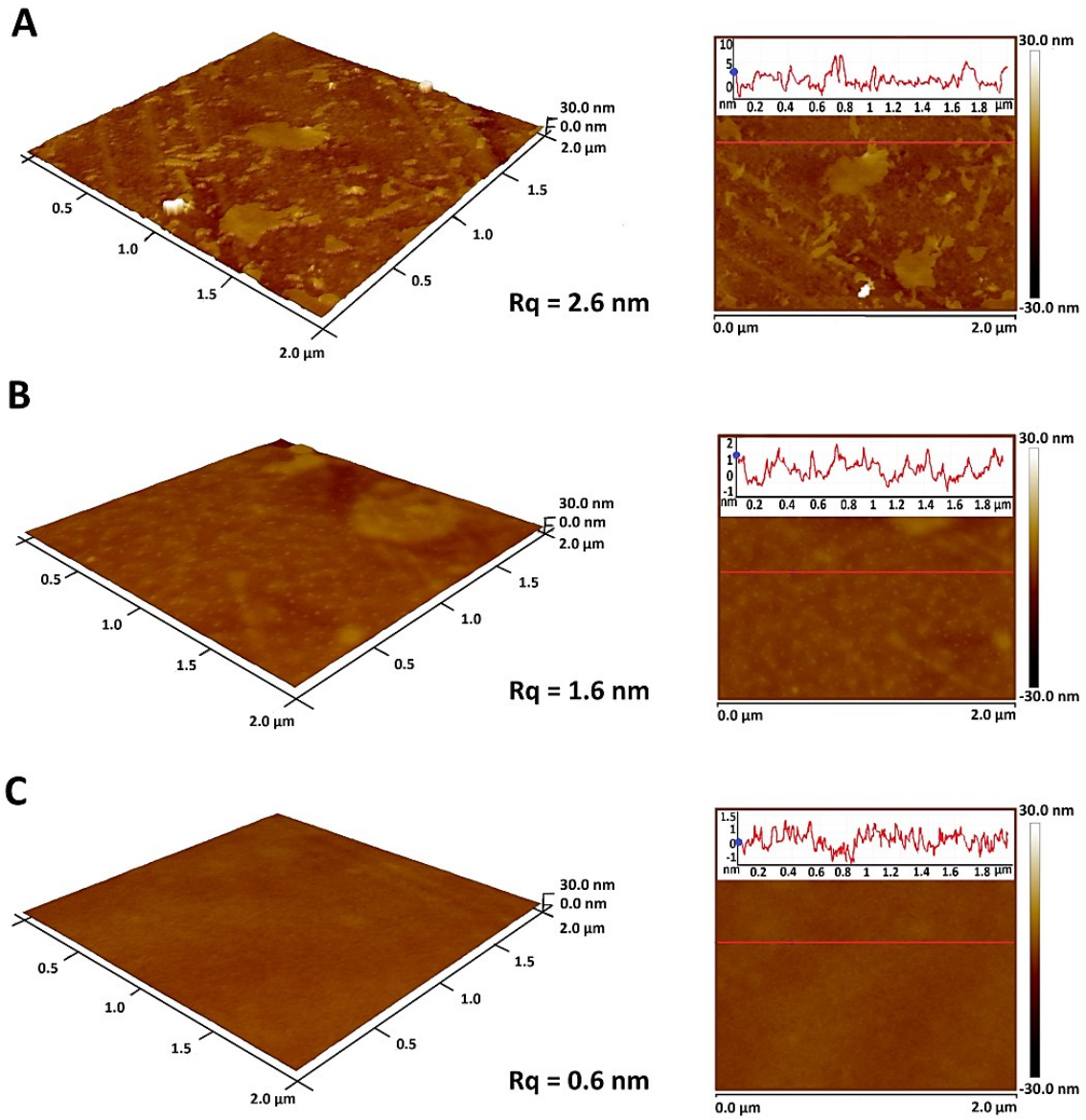


Figure 8: Atomic force microscopy representations of the surface topography of samples: A) untreated PET film, B) ethyl lactate plasma-treated sample (flow rate 1000 sccm), C) methyl lactate plasma-treated sample (flow rate 1000 sccm).

### 3.4 Antibacterial properties of the tested materials

The antibacterial activity of the prepared films was evaluated in accordance with the international standard ISO 22196:2011 [15]. The assessment was conducted on sample films containing methyl lactate and ethyl lactate, using *E. coli* as a representative gram-negative

bacterial strain and *S. aureus* as an example of gram-positive strain. As can be seen in Table 3, samples with methyl lactate exhibit greater antimicrobial efficacy. Notably, sample 6, deposited at a flow rate of 500 sccm, demonstrated strong antibacterial activity (R=3.4) against *E. coli* and significant activity (R=2.1) against *S. aureus*. Furthermore, sample 5 has been found to be particularly relevant, as it exhibits significant antimicrobial properties against both tested bacterial strains.

This antibacterial activity may be associated with the presence of amines, as suggested by XPS analysis. Several studies have confirmed that amines possess notable antibacterial properties [37–39]. Mentheour et al. reported that the antibacterial effects were primarily linked to enhanced hydrophilicity resulting from amines generated by reactive nitrogen species (RNS) within the dielectric barrier discharge (DBD) plasma source [40].

In general, surfaces exhibiting moderate wettability tend to facilitate bacterial or cellular adhesion more effectively than surfaces that are either highly hydrophobic or extremely hydrophilic. Additionally, an increase in surface free energy (Table 2), particularly in its polar or acid-base components, along with an enhancement of the electron donor character of the substrate, has been shown to reduce bacterial adhesion [36,41]. The bacterial-repellent properties of hydrophilic surfaces can be attributed to the formation of a hydration layer on PET-treated surfaces. This hydration layer, arising from the hydrophilic and polar nature of the surface, generates hydration forces that inhibit the attachment of non-specific proteins and microorganisms [42].

Surface roughness is another critical factor that can inhibit bacterial adhesion by increasing the available surface area for cell attachment and reducing the shear force of the flowing fluid. This reduction in shear force helps prevent the detachment of adhered bacteria [41]. However, nanoscale surface roughness has been found to exhibit the most effective anti-adhesion properties, whereas microscale roughness tends to promote bacterial adhesion. In contrast, smooth surfaces influence bacterial behaviour primarily through other material properties, such as surface chemistry and surface energy [36].

It should be noted that the long-term stability of the antibacterial properties was not evaluated in the present study and will be the subject of future investigation.

Table 3 Antibacterial activity (R) of films containing methyl lactate and ethyl lactate, deposited at varying monomer flow rates, against gram-positive and gram-negative bacterial strains after 24 hours of incubation.

| Samples | *Staphylococcus aureus* CCM 4516 | | *Escherichia coli* CCM 4517 | |
|---|---|---|---|---|
| | **$N_x$(cfu/cm$^2$)** | **R** | **$N_x$(cfu/cm$^2$)** | **R** |
| 0. PET reference | 2.4x10$^5$ | $U_t$ = 5.4 | 1.6x10$^6$ | $U_t$ = 6.2 |
| 1. ethyl lactate/PET | 1.0x10$^5$ | 0.4 | 4.3x10$^5$ | 0.6 |
| 2. ethyl lactate/PET | 2.6x10$^4$ | 1.0 | 2.9x10$^5$ | 0.7 |
| 3. ethyl lactate/PET | 6.8x10$^4$ | 0.5 | 1.5x10$^5$ | 1.1 |
| 4. ethyl lactate/PET | 3.1x10$^4$ | 0.9 | 3.9x10$^5$ | 0.6 |
| 5. methyl lactate/PET | 1.2x10$^3$ | 2.3 | 1.7x10$^4$ | 2.0 |
| 6. methyl lactate/PET | 1.9x10$^3$ | 2.1 | 6.5x10$^2$ | 3.4 |
| 7. methyl lactate/PET | 6.1x10$^4$ | 0.6 | 4.2x10$^5$ | 0.6 |
| 8. methyl lactate/PET | 9.2x10$^4$ | 0.4 | 2.8x10$^5$ | 0.8 |

## 4 Conclusion

In this study, lactate esters were obtained through the environmentally sustainable chemical recycling of a biopolymer and subsequently modified via plasma deposition. This process enabled the development of a method for depositing antibacterial films on PET surfaces.

ATR-FTIR spectroscopy confirmed the presence of O–H bonds associated with the enhanced hydrophilicity of the samples, particularly in the ethyl lactate-based coating (sample 3). Complementary insights were provided by XPS analysis, which revealed the formation of oxygen- and nitrogen-containing functional groups, with C=O, C–N, and N–C=O bonds being most pronounced in the ethyl lactate/PET sample. Considering that XPS probes only the top ~5–10 nm of the surface, whereas ATR-FTIR provides information from depths up to the micrometer scale, the combined use of these techniques enables a comprehensive evaluation of both surface-specific and bulk-like chemical characteristics. Although the film thickness was not measured directly, it is reasonable to assume that it falls within the range of tens to hundreds of nanometers, as reported based on comparable atmospheric-pressure plasma polymerization systems.

The chemical composition and surface morphology analyses revealed significant differences in topography between plasma-treated and untreated samples. Notably, the plasma-treated films exhibited antibacterial properties against both *S. aureus* and *E. coli* in comparison to

untreated PET film. This antibacterial effect may be attributed to the hydrophilicity of the coatings and the presence of amines.

The plasma deposition of degradation products derived from the chemical recycling of biopolymers presents a novel approach to sustainable material development within the framework of the circular economy. This method not only introduces materials with enhanced functional properties but also offers an innovative strategy for sustainable waste management by utilizing chemically recycled polylactic acid esters.

## CRediT authorship contribution statement

Monika Strašáková: Writing – original draft, Visualization, Validation, Methodology, Investigation, Data curation, Conceptualization. Eva Domincová Bergerová: Project administration, Methodology, Investigation, Conceptualization, Marián Lehocký: Methodology, Investigation. Veronika Hanuliak: Methodology, Investigation. Hana Pištěková: Methodology, Investigation. Michal Urbánek: Methodology, Investigation. Pavel Šťahel: Methodology, Investigation. Monika Stupavská: Methodology, Investigation. Jakub Klaban: Methodology, Investigation. Anežka Lengálová: Writing – review & editing. Vladimír Sedlařík: Supervision, Funding acquisition, Formal analysis.

## Declaration of competing interest

The authors declare that they have no conflict of interest.

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## Data availability

Data will be made available on request.

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